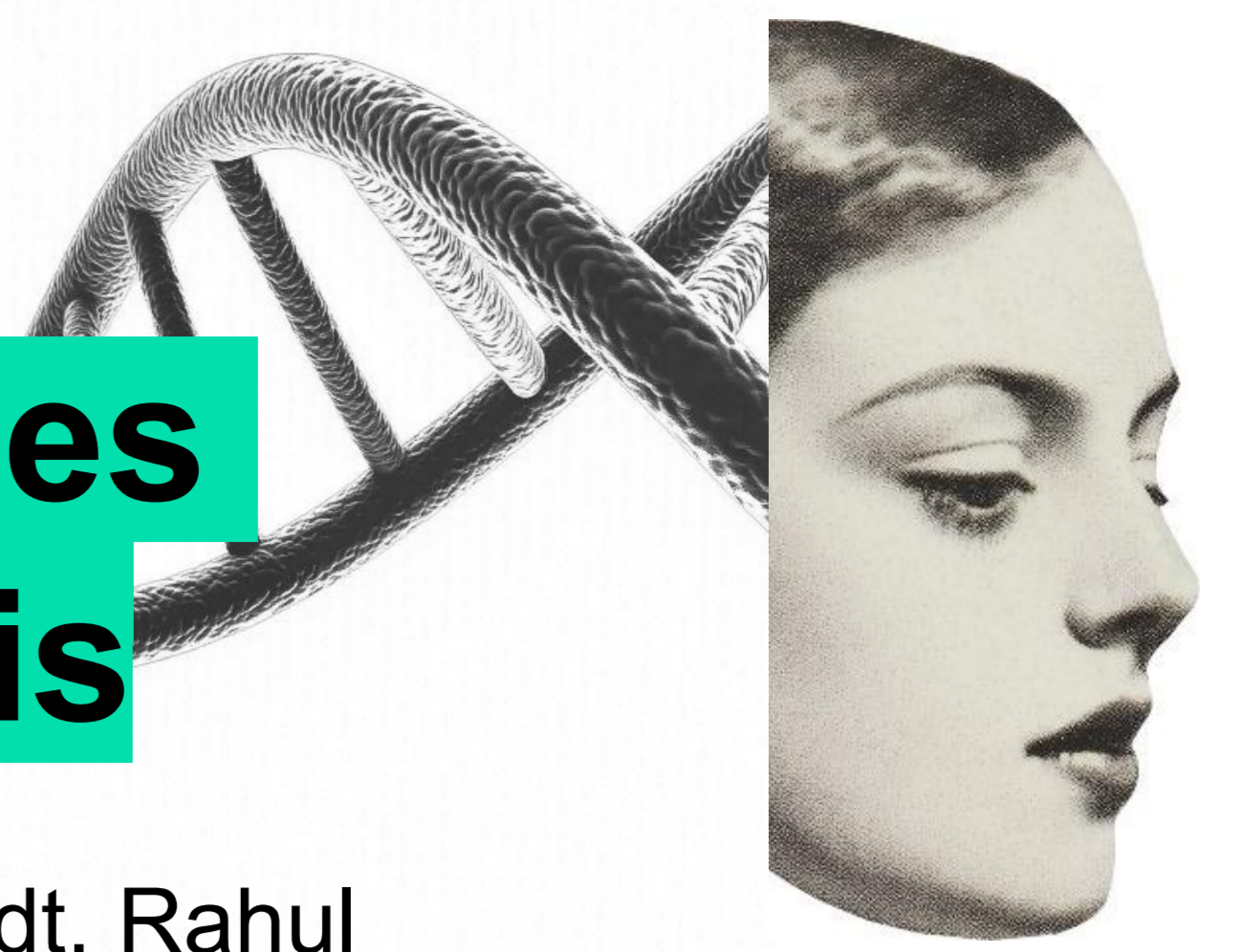




Design of highly efficient sgRNA libraries through comprehensive feature analysis

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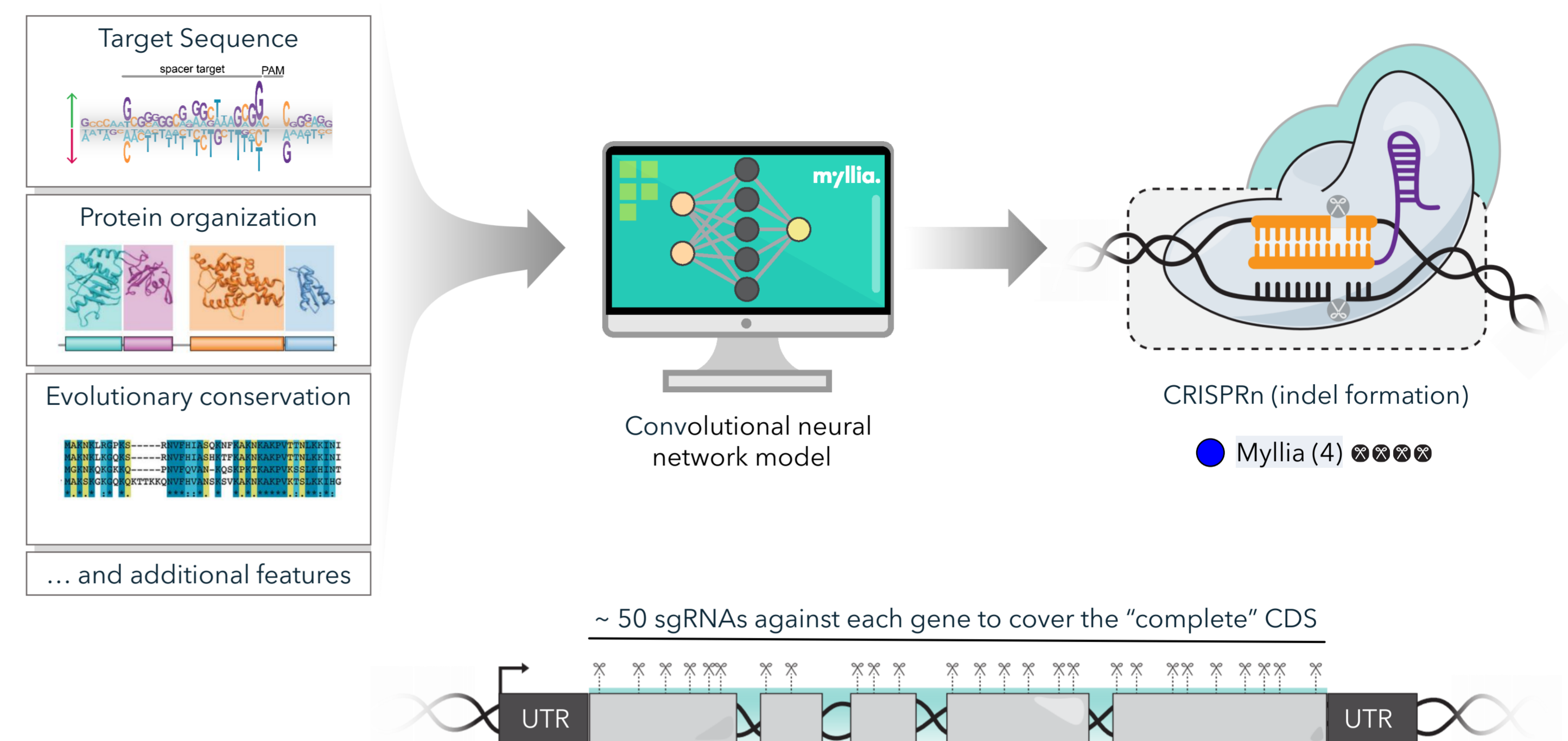
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About Myllia Biotechnology

Myllia Biotechnology combines CRISPR screening with single-cell RNA sequencing, leveraging two powerful technologies to enable detailed analysis of complex phenotypes. Using CROP-Seq alongside other high-content screening technologies, we systematically investigate the effects of thousands of genetic perturbations in primary human cells. Our platform delivers a comprehensive view of transcriptomic changes at single-cell resolution, revealing how genetic perturbations shape cellular behaviour and disease mechanisms. This innovative approach supports a wide range of applications, including the discovery of novel drug targets, elucidating the mechanism of action of drugs, and understanding genetic variants linked to disease risk.

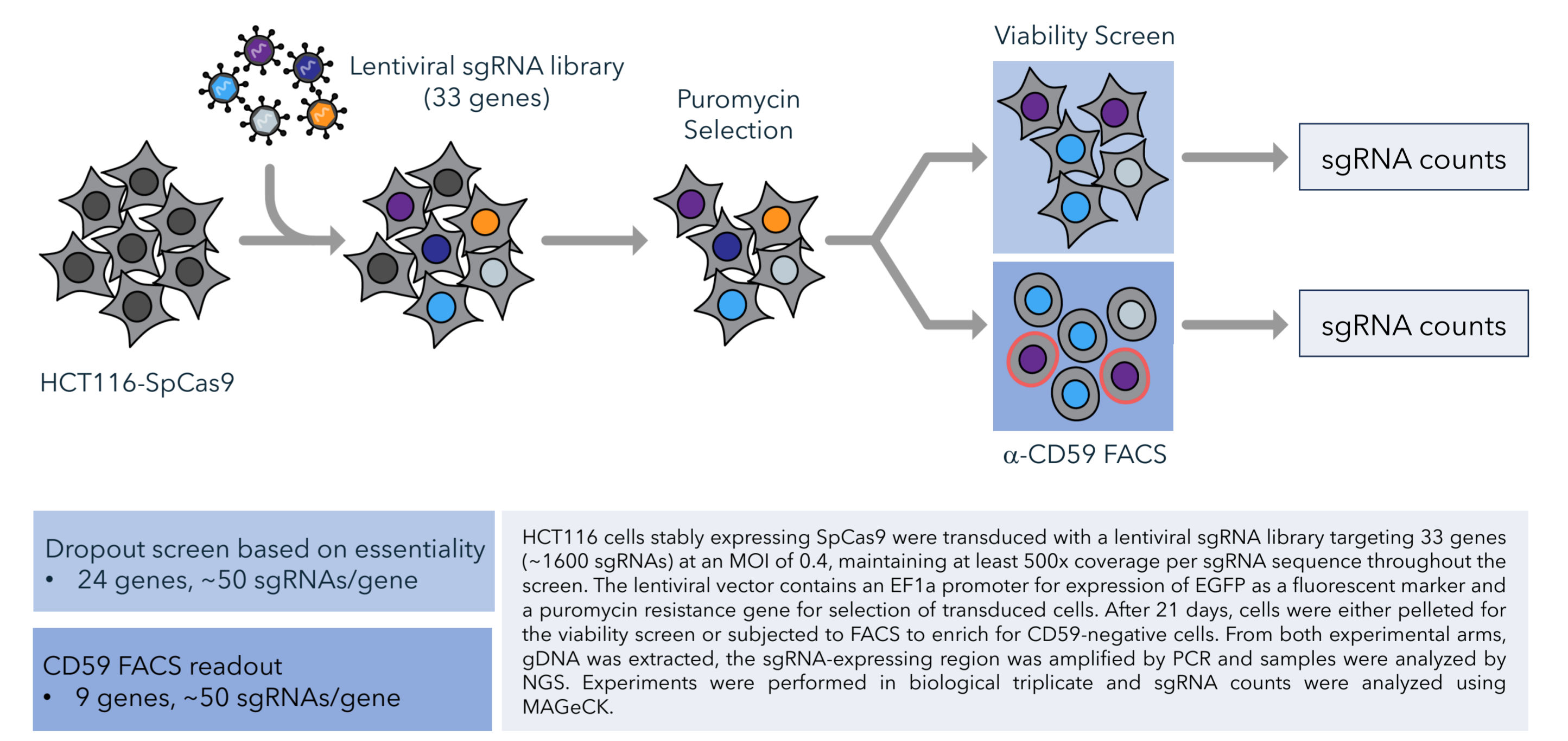
1 The sgRNA design algorithm for CRISPRn screens



2 Commonly used genome-scale sgRNA libraries

<p>GeCKO_v2 </p> <p>Correspondence Published: 30 July 2014</p> <p>Improved vectors and genome-wide libraries for CRISPR screening</p> <p><i>Neville E Sanjana, Qibin Shalem & Feng Zhang</i></p>	<p>Behan </p> <p>Prioritization of cancer therapeutic targets using CRISPR-Cas9 screens</p> <p><i>Fiona M. Behan^{1,2,3}, Francesco Picco^{1,2,3}, Gabriele Piccini^{1,2}, Immanuel Gschwend¹, Charlotte M. Beaver¹, Caterina Migliardi^{1,4}, Silia Ballester¹, Yanhui Bao¹, Francesco Sassi¹, Maria Pavesi^{1,5}, Roman Kozlov¹, Sarah Harper¹, David Adam Jackson¹, Rebecca McKee¹, Rachel Pook¹, Peter Wilkerson¹, Dorothea van der Meer¹, David Dowse¹, Candice Bauer¹, Deepankar¹, Andrea Bernhart¹, Liron Trudler¹, Susan A. Strassman^{1,6}, Julio Saez-Rodriguez^{1,7,8,9}, Kunal Talwar^{1,10,11} & Matthew J. Garnett^{1,12}</i></p>
<p>TKO_v3 </p> <p>Evaluation and Design of Genome-Wide CRISPR/SpCas9 Knockout Screens</p> <p><i>Traver Hart^{1*}, Amy Hui Yan Tong¹, Katie Chan¹, Jelanda Van Leeuwen¹, Adhwin Soetharaman¹, Michael Angger¹, Megha Chandrasekhar¹, Nicole Hustedt¹, Sahil Sethi¹, Avery Noonan¹, Andrea Habsid¹, Olga Sizova¹, Lyudmila Nedyalkova¹, Ryan Climie¹, Leanne Tworzynski¹, Keith Lawson¹, Maria Augusta Sartori¹, Sabriyah Alibek¹, David Tieu¹, Sanna Masud¹, Patricia Mero¹, Alexander Weiss¹, Kevin R. Brown¹, Matej Uraj¹, Maximilian Billmann¹, Mahfuzur Rahman¹, Michael Costanzo¹, Chad L. Myers¹, Brenda J. Andrews^{1,2,3}, Charles Boone^{1,4,5}, Daniel Durocher^{1,6} and Jason Moffat^{1,7,8,9}</i></p>	<p>VBC </p> <p>Multilayered VBC score predicts sgRNAs that efficiently generate loss-of-function alleles</p> <p><i>Georg Michlits^{1,2}, Julian Jude^{1,2,3}, Matthias Hinterndorfer², Melanie de Almeida^{1,2}, Gintautas Vainorius¹, Maria Hubmann¹, Tobias Neumann^{1,2}, Alexander Schlieffer^{1,2}, Thomas Rainer Burkard^{1,2}, Michaela Fellner¹, Max Gijbetsen¹, Anna Traunbauer¹, Johannes Zuber^{1,2,3,4} and Ulrich Elling^{1,2,3}</i></p>
<p>Brunello </p> <p>Optimized libraries for CRISPR-Cas9 genetic screens with multiple modalities</p> <p><i>Kendall R. Sanson¹, Ruth E. Hanna¹, Mudra Hegde¹, Katherine F. Donovan¹, Christine Strand¹, Meagan E. Sullender¹, Emma W. Vaimberg¹, Amy Goodale¹, David E. Root¹, Federica Piccioni¹ & John G. Doench¹</i></p>	<p>MinLibCas9 </p> <p>Minimal genome-wide human CRISPR-Cas9 library</p> <p><i>Emanuel Gonçalves¹, Mark Thomas¹, Fiona M. Behan¹, Gabriele Piccini¹, Clare Pacini^{1,2}, Felicity Allen¹, Alessandro Vinceti¹, Mamta Sharma¹, David A. Jackson¹, Stacy Price¹, Charlotte M. Beaver¹, Oliver Dovey¹, David Pary-Smith¹, Francesco Iorio^{1,3}, Leopold Parts^{1,4}, Kosuke Yusa¹ and Matthew J. Garnett^{1,5}</i></p>

3 Screens to study essential and non-essential genes

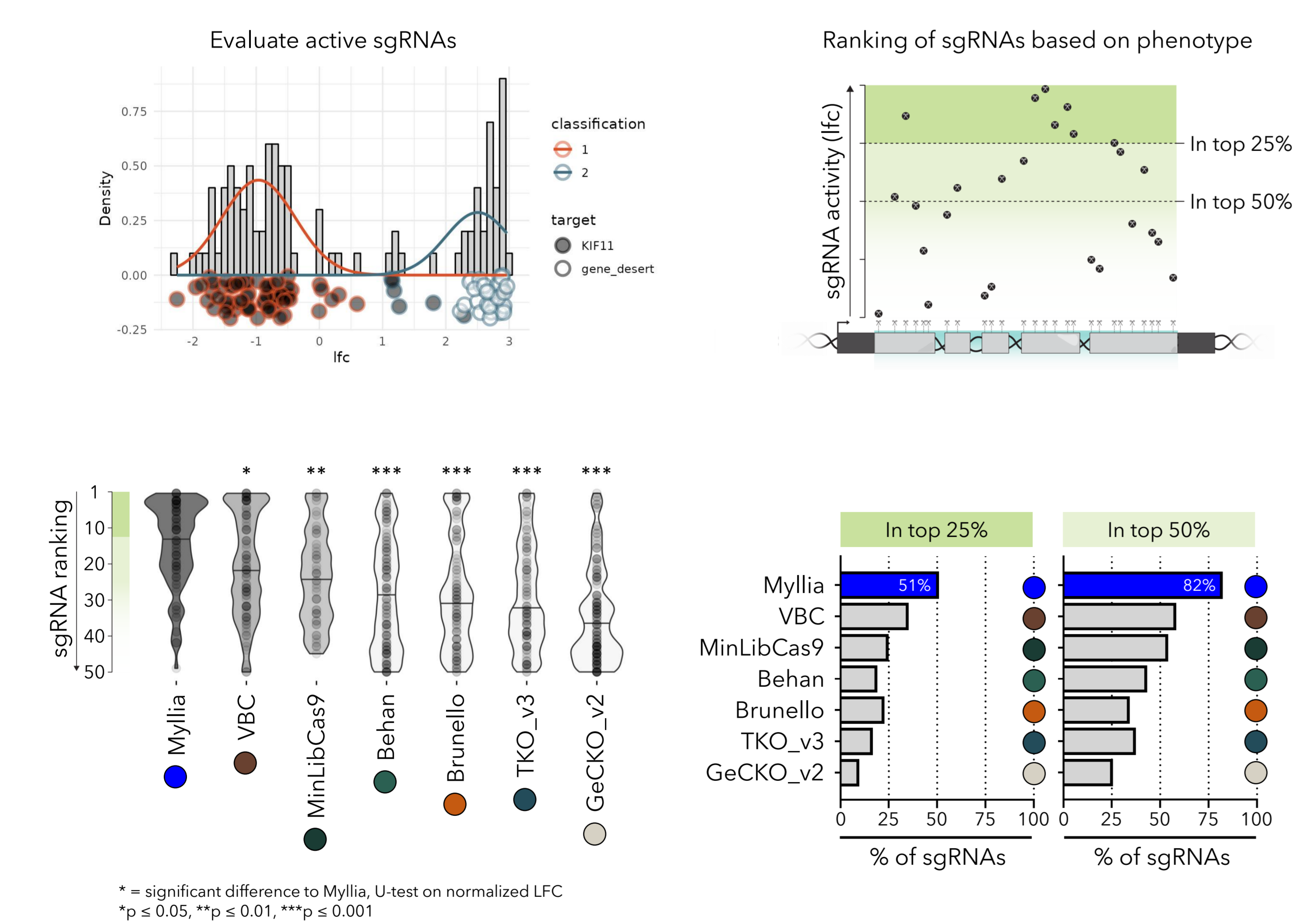


Dropout screen based on essentiality
 • 24 genes, ~50 sgRNAs/gene

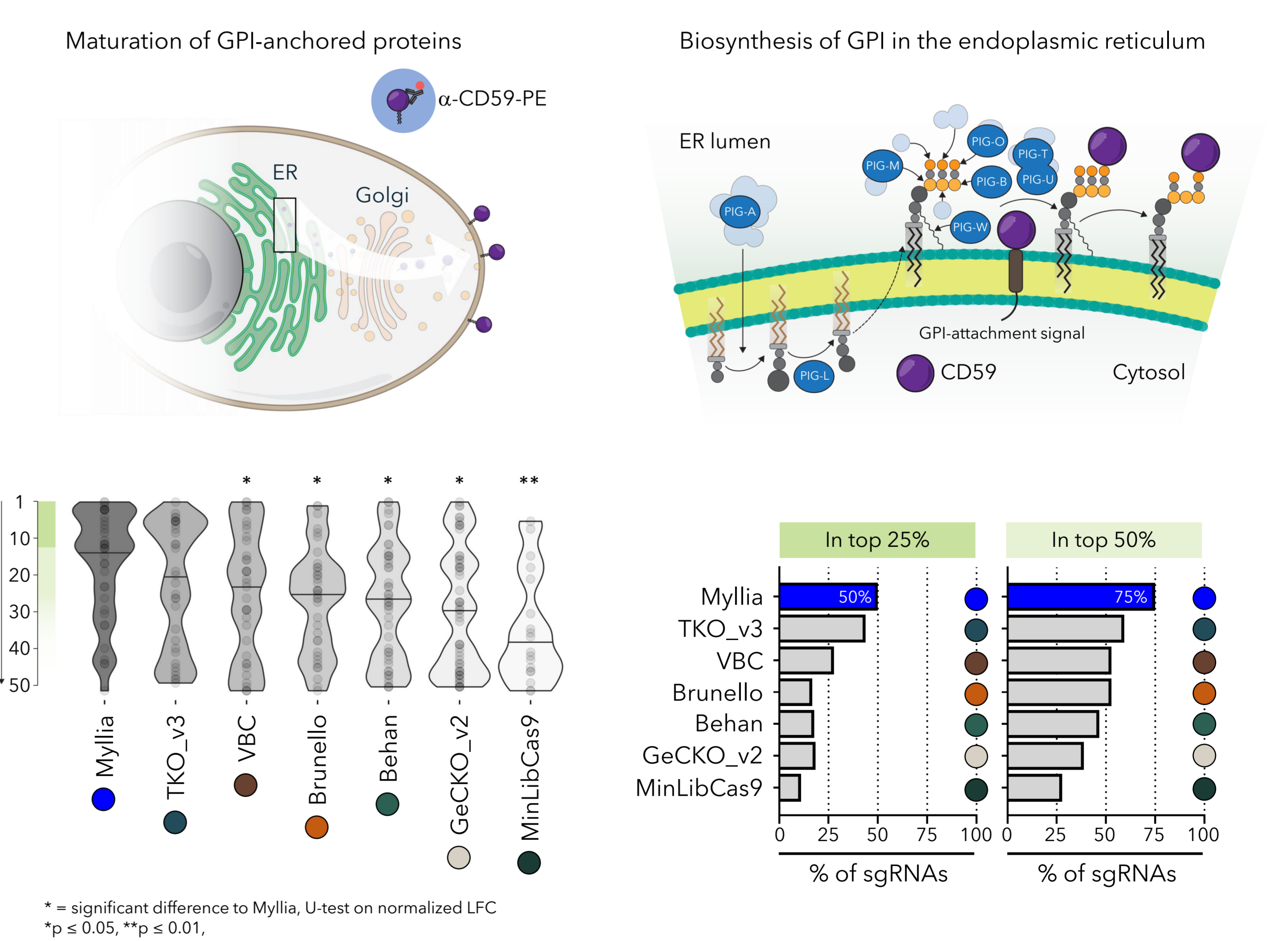
CD59 FACS readout
 • 9 genes, ~50 sgRNAs/gene

HCT116 cells stably expressing SpCas9 were transduced with a lentiviral sgRNA library targeting 33 genes (~1600 sgRNAs) at an MOI of 0.4, maintaining at least 500x coverage per sgRNA sequence throughout the screen. The lentiviral vector contains an EF1a promoter for expression of EGFP as a fluorescent marker and a puromycin resistance gene for selection of transduced cells. After 21 days, cells were either pelleted for the viability screen or subjected to FACS to enrich for CD59-negative cells. From both experimental arms, gDNA was extracted, the sgRNA-expressing region was amplified by PCR and samples were analyzed by NGS. Experiments were performed in biological triplicate and sgRNA counts were analyzed using MAGeCK.

4 Viability phenotype to assess sgRNA activities



5 FACS-based screen to monitor CD59 localization



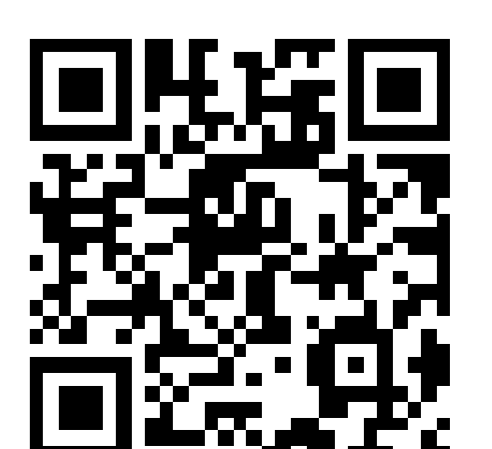
Conclusions and Outlook

- Myllia's sgRNA design algorithm selects highly active sgRNA sequences
- Our sgRNA libraries contain optimal sgRNA sequences for both targeting essential and non-essential genes
- We will expand the screening campaign to target additional non-essential genes that could help to further improve sgRNA designs and increase editing rates
- CRISPRi libraries will be evaluated using a comparable workflow



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